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Chasing after Non-cyanobacterial Nitrogen Fixation in Marine Pelagic Environments

Pia H. Moisander^{1*}, Mar Benavides², Sophie Bonnet³, Ilana Berman-Frank⁴, Angelique E. White⁵ and Lasse Riemann²

¹ Department of Biology, University of Massachusetts Dartmouth, North Dartmouth, MA, United States, ² Marine Biology Section, Department of Biology, University of Copenhagen, Helsingør, Denmark, ³ Centre National de la Recherche Scientifique, IRD, Aix-Marseille Université, Université de Toulon, Marseille, France, ⁴ Mina and Everard Goodman Faculty of Life Sciences, Bar-Ilan University, Ramat Gan, Israel, ⁵ College of Earth, Ocean, and Atmospheric Sciences, Oregon State University, Corvallis, OR, United States

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*Correspondence:

Pia H. Moisander
pmoisander@umassd.edu

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Traditionally, cyanobacterial activity in oceanic photic layers was considered responsible for the marine pelagic dinitrogen (N₂) fixation. Other potentially N₂-fixing bacteria and archaea have also been detected in the pelagic water column, however, the activity and importance of these non-cyanobacterial diazotrophs (NCDs) remain poorly constrained. In this perspective we summarize the N₂ fixation rates from recently published studies on photic and aphotic layers that have been attributed to NCD activity via parallel molecular measurements, and discuss the status, challenges, and data gaps in estimating non-cyanobacterial N₂ fixation NCNF in the ocean. Rates attributed to NCNF have generally been near the detection limit thus far (<1 nmol N L⁻¹ d⁻¹). Yet, if considering the large volume of the dark ocean, even low rates of NCNF could make a significant contribution to the new nitrogen input to the ocean. The synthesis here shows that *nifH* transcription data for NCDs have been reported in only a few studies where N₂ fixation rates were detected in the absence of diazotrophic cyanobacteria. In addition, high apparent diversity and regional variability in the NCDs complicate investigations of these communities. Future studies should focus on further investigating impacts of environmental drivers including oxygen, dissolved organic matter, and dissolved inorganic nitrogen on NCNF. Describing the ecology of NCDs and accurately measuring NCNF rates, are critical for a future evaluation of the contribution of NCNF to the marine nitrogen budget.

Keywords: bacteria, diazotroph, DOM, mesopelagic, *nifH*, nitrogenase, oxygen minimum zone

INTRODUCTION

Biological dinitrogen (N₂) fixation produces biologically available nitrogen (N) through reduction of atmospheric N₂ to ammonium (NH₄⁺) (Postgate, 1998). In oligotrophic marine environments, N₂ fixation can provide ~50% of the “new” N (Karl et al., 2002; Capone et al., 2005; Berthelot et al., 2017), and probably contributes more in hot spots such as the Western Tropical South Pacific (Bonnet et al., 2017). N₂ fixation (diazotrophy) is catalyzed by the enzyme nitrogenase (Mortenson and Thorneley, 1979; Postgate, 1998). The *nifH* gene that encodes the dinitrogenase reductase is distributed in many phylogenetic groups of bacteria and archaea (Chien and Zinder, 1996), and used to assess the diversity and expression of the enzyme in marine diazotrophs (Zehr et al., 2003).

Cyanobacteria have traditionally been considered the most important diazotrophs in the ocean. The predominantly described taxa include the filamentous, bloom-forming *Trichodesmium*, heterocystous, symbiotic groups (e.g., diatom-diazotroph associations), and the unicellular cyanobacteria UCYN-A (*Candidatus Atelocyanobacterium thalassa*), UCYN-B (*Crocospaera watsonii*), and UCYN-C (Zehr, 2011). Early molecular studies also reported the presence of non-cyanobacterial diazotrophs (NCDs) (Kirshtein et al., 1991; Zehr et al., 1995, 1998). These findings stimulated recent research on diversity, composition, and ecology of marine NCDs (Bombar et al., 2016), leading to a currently perceived emergence of a new paradigm: non-cyanobacterial N₂ fixation (NCNF). Extensive research on free-living and symbiotic NCDs in soils has provided evidence of the activity, regulation, and the significance of these processes in terrestrial ecosystems (Postgate, 1998; Herridge et al., 2008; Hayat et al., 2010). While the diversity and abundance of marine NCDs indicate that NCNF could potentially have a large impact on the marine N budget, at present, the activity and contribution of marine NCNF to total N₂ fixation remain poorly constrained.

In many oceanic waters, NCD sequences dominate the *nifH* gene pool (Riemann et al., 2010; Farnelid et al., 2011), and detection of transcripts of some of the NCD *nifH* genes suggests that at least some of the NCDs fix N₂ in the oceanic water column and sediments (Bird et al., 2005; Church et al., 2005b; Halm et al., 2012; Brown and Jenkins, 2014; Moisander et al., 2014; Bentzon-Tilia et al., 2015b). However, gene expression cannot be equated with active N₂ fixation rates without evidence of cell-specific activity. Current broadly available methods cannot discern the relative contribution of cyanobacteria and NCDs to measured N₂ fixation rates. The contribution of NCNF could be studied by parallel rate measurements and molecular detection in areas where cyanobacterial diazotrophs are typically not present (such as the oceanic water column below the euphotic layer, i.e., aphotic waters). Here we summarize data from studies where N₂ fixation rates were reported in parallel to molecular characterization of NCDs when cyanobacterial diazotrophs were *not* detected (presence and/or expression of the *nifH* gene; **Table 1**). Presumed NCNF rates ranged from undetectable to 8 and 0.89 nmol N L⁻¹ d⁻¹ in photic and aphotic studies, respectively (**Table 1**). The goal of this perspective is to synthesize emerging trends and data gaps from these studies and highlight future research directions.

NCNF IN PHOTIC WATERS

Most early studies of oceanic N₂ fixation (Montoya et al., 1996; Karl et al., 2002; Hood et al., 2004; Mahaffey et al., 2005; Mulholland et al., 2006) and diazotroph diversity and activity (Zehr, 2011) focused on photic waters and cyanobacterial diazotrophs. Traditionally, the photic layer has been considered an optimal environment selecting for cyanobacterial N₂ fixation, as abundant light energy (harnessed via photosynthesis) supplies the high energetic demands of the N₂ fixation process, while dissolved inorganic nitrogen (DIN) limits non-diazotrophic

autotrophs. Yet, since the first molecular studies (Zehr et al., 1998), *nifH* sequence libraries have revealed diverse NCD phylotypes from surface waters of various oceans and estuaries (Falcon et al., 2004; Church et al., 2005a; Langlois et al., 2005; Hewson et al., 2007; Moisander et al., 2008; Foster et al., 2009; Farnelid et al., 2011; Halm et al., 2012; Bentzon-Tilia et al., 2015b; Messer et al., 2016). Transcripts from a range of NCD groups have been recovered in surface layers (Man-Aharonovich et al., 2007; Halm et al., 2012; Loescher et al., 2014). One of the few consistent NCDs in photic layers is a gamma-proteobacterial cluster of sequences (termed γ -24774A11, Gamma A, or UMB; representatives of the same phylotype). Gamma A shows a wide distribution and expression of the *nifH* gene (Bird et al., 2005; Church et al., 2005b; Moisander et al., 2014; Langlois et al., 2015). This group appears to be broadly distributed across tropical and subtropical surface waters compared with cyanobacterial diazotrophs (Moisander et al., 2014; Bonnet et al., 2015; Langlois et al., 2015). Its presence exclusively in surface layers suggests either reliance on a photosynthetic machinery, rhodopsin, or photosynthesis products from other organisms. Once cultivated or its genome assembled, we can learn more about its autecology. A few other studies have detected N₂ fixation from photic layers in the reported absence of cyanobacterial sequences (Yogev et al., 2011; Blais et al., 2012). However, due to the difficulty of demonstrating the absence of cyanobacterial diazotrophs, it has generally been difficult to prove that NCNF is active in photic layers.

The nitrogenase enzyme may be downregulated or inactivated by NH₄⁺ (Zehr et al., 1997). As typically assumed for cyanobacterial diazotrophs (Zehr, 2011), DIN could negatively impact NCDs directly (physiological inhibition) or indirectly (NCDs outcompeted by faster growing non-diazotrophs). However, DIN availability may not always have immediate negative effects on NCDs. For example, Gamma A transcripts and Cluster III diazotrophs (Chien and Zinder, 1996) were found in the presence of micromolar concentrations of DIN in the upper layers of the Arabian Sea (Bird and Wyman, 2013). N₂ fixation rates were also detected in the Benguela Upwelling System in the presence of micromolar nitrate and in the absence of cyanobacterial diazotrophs (Sohm et al., 2011). In other surface waters (Eastern Tropical South Pacific; ETSP), N₂ fixation rates were similarly reported in the absence of cyanobacterial diazotrophs and presence of high DIN (Fernandez et al., 2011; Dekaezemacker et al., 2013; Gradoville et al., 2017). In Indian Ocean surface waters with a shallow nitracline, N₂ fixation rates were reported in parallel with up to 10⁴ *nifH* gene copies L⁻¹ of Gamma A, while cyanobacterial groups were undetectable (Shiozaki et al., 2014). Collectively these field studies suggests that NCDs may have a low sensitivity to DIN (Knapp, 2012), but recent work with cultivated NCDs suggest that responses to DIN may be strain specific (Bentzon-Tilia et al., 2015a).

PCR amplification biases likely influence our current data and conclusions of studies on NCDs. A recent study using metagenomic data from the TARA Oceans survey reports

TABLE 1 | Compilation of photic and aphotic N₂ fixation rates published with molecular data and associated with non-cyanobacterial diazotrophs in waters reportedly devoid of cyanobacterial diazotrophs.

References	Location	Latitude	Longitude	Average rate or range of rates (nmol N L ⁻¹ d ⁻¹)	Depth (m)	Sequencing: Associated non-cyanobacterial phylogeny genes and transcripts	qPCR: non-cyanobacterial diazotroph gene and transcript abundance (<i>nifH</i> copies L ⁻¹)
SOUTH AND NORTH PACIFIC OCEAN							
Benavides et al., 2015	Western South Pacific	3–9°S	146–153°E	0.08–0.89	200–1000	Cluster I, III, IV, and <i>Trichodesmium</i> nd	0–900 nd
Halm et al., 2012	South Pacific Gyre	20–50°S	180°E–120°W	0.41.5	0–200	Cluster I, III nd	0–7*10 ⁶ 0–8*10 ⁴
Fernandez et al., 2011	Eastern Tropical South Pacific	1.5°N–17.8°S	86.9–70.8°W	0.01–3.5	70–400 ^a	Cluster I, II, III nd	nd nd
Dekazemackner et al., 2013	Eastern Tropical South Pacific	10–20°S	100–80°W	0–0.88	15–150	b b	b b
Turk-Kubo et al., 2014	Eastern Tropical South Pacific	10–20°S	100–80°W	b	10–200	Cluster I, III Cluster I	0–420 ~400
Loescher et al., 2014	Eastern Tropical South Pacific	2°N–16°S	85–74°W	0.4	0–350	Cluster I, III, IV nd	0–10 ⁶ 0–150
Bonnet et al., 2013	Eastern Tropical South Pacific	10–20°S	100–80°W	0–0.8	200–2,000	Cluster I, III nd	nd nd
Gradoville et al., 2017	Eastern Tropical South Pacific	20.1–26.3°S	104–77°W	0.5–5.1	5 m for N ₂ fix, 5–420 m for <i>nifH</i>	Cluster I, III nd	1.5 × 10 ² –1.4 × 10 ⁴ nd
Hamersley et al., 2011	Southern California Bight	33.55°N	118.4°W	0.07	500–885	Cluster I, III, IV nd	nd nd
ATLANTIC OCEAN, ARCTIC OCEAN							
Sohm et al., 2011	South Atlantic Gyre and Benguela upwelling system	11–25°S	29°W–15°E	0.06–8	8 m for N ₂ fix, 8–110 m for <i>nifH</i>	nd nd	nd nd
Blais et al., 2012	Arctic Ocean coastal	69.3–75°N	69–134°W	0.02–4.45	5 m, DCM (30–57)	Cluster I, III nd	nd nd
INDIAN OCEAN							
Shiozaki et al., 2014	Indian Ocean	4°N–20°S	65–70°E	0.18–0.23	0–90	Cluster I nd	10 ³ nd

(Continued)

TABLE 1 | Continued

References	Location	Latitude	Longitude	Average rate or range of rates (nmol N L ⁻¹ d ⁻¹)	Depth (m)	Sequencing: Associated non-cyanobacterial phylotype genes and transcripts	qPCR: non-cyanobacterial diazotroph gene and transcript abundance (<i>nifH</i> copies L ⁻¹)
Jayakumar et al., 2012	Indian Ocean	13–19°N	64°–66°E	nd	110–175 (OMZ)	Cluster I, III, IV Cluster I, III	nd nd
MEDITERRANEAN SEA, RED SEA, AND BALTIC SEA							
Rahav et al., 2013a	Mediterranean Sea, Red Sea	29°55N–32°57N	34°29E–34°45E	0.01–0.38	150–720	Pseudomonas stutzeri-related nd	nd nd
Rahav et al., 2016	Eastern Mediterranean Sea	32°N	34°E	0.1–0.15	5	nd Cluster I: (Alpha-), and “other”	nd nd
Yogev et al., 2011	Eastern Mediterranean Sea	33.14–34.00°N	25°–33°E	0–0.3	0–160m	Cluster I, II Cluster I, II	nd nd
Farnelid et al., 2013	Baltic Sea	57.20°N	20.03°E	0.44	200	Cluster I, II, III ^d Cluster I, III	0–2*10 ⁷ 0–10 ⁴

All rates are based on the ¹⁵N₂ method and no conversions were made to originally reported values. The station location and depth information is specific for locations where **no** cyanobacterial diazotrophs were detected (unless otherwise indicated). Red letters indicate transcripts. Clusters I, II, III, and IV follow Chien and Zinder (1996).

^aDepths range from photic to aphotic.

^bDekaezemacker et al. (2013) reported rates and Turk-Kubo et al. (2014) reported corresponding molecular data.

^cnd, data not available/not done.

^dNodularia transcripts were detected at abundances below the level of quantification via RT-qPCR and its transcripts were not detected by sequencing.

presence of nitrogenase containing Planctomycetes in oceanic photic waters and suggests they have a substantially greater abundance than the NCDs reported based on past PCR approaches (Delmont et al., 2017).

NCNF IN APHOTIC WATERS AND OXYGEN DEFICIENT ZONES

Several studies have recently reported N₂ fixation rates in the aphotic ocean where active autotrophic cyanobacterial diazotrophy is not expected. Early evidence for aphotic N₂ fixation showed proteobacterial and archaeal diazotrophs and N₂ fixation in hydrothermal vent fluids (Mehta and Baross, 2006). Deep-sea sediment archaea fixing N₂ apparently function in associations with anaerobic methane oxidizers (Dekas et al., 2009, 2014). Proteobacterial *nifH* sequences were reported from the meso- to abyssopelagic water column in different oceans (Hewson et al., 2007; Moisander et al., 2008).

Laboratory studies have shown that nitrogenase activity in cells is reduced under increasing concentrations of oxygen, and the enzyme is irreversibly modified by oxygen *in vitro* (Fay, 1992; Berman-Frank et al., 2008). Thus, oxygen deficient zones (ODZs) that may span photic and aphotic layers could serve as sites of N₂ fixation due to low oxygen, in parallel with N losses from these zones (Deutsch et al., 2007). The first studies reporting planktonic N₂ fixation rates in aphotic waters were ones associated with ODZs in the Eastern Tropical North Pacific Ocean (ETNP) (Hamersley et al., 2011) and ETSP (Fernandez et al., 2011; Dekaezemacker et al., 2013). N₂ fixation rates were reported in mesopelagic waters down to 400 and 800 m, and abyssopelagic waters down to 2,000 m (Fernandez et al., 2011; Bonnet et al., 2013; Loescher et al., 2014). These rates were attributed to various NCD DNA sequences, including Cluster I (Proteobacteria), II and III (Turk-Kubo et al., 2014). N₂ fixation in the presence of NCD *nifH* expression was also reported in the aphotic ODZ of the Baltic Sea (Farnelid et al., 2013), and in the ODZ of the Arabian Sea (Jayakumar et al., 2012). These studies suggest that NCNF may be active in ODZs, however evidence of the relationship between oxygen and NCNF is variable. N₂ fixation in the aphotic Western Tropical South Pacific had a negative correlation with dissolved oxygen (Benavides et al., 2015). However, mesopelagic N₂ fixation rates in the Mediterranean Sea were reported from fully oxygenated waters (Rahav et al., 2013a), or were negatively correlated with apparent oxygen utilization values (i.e., N₂ fixation was higher in more recently ventilated waters; Benavides et al., 2016). Gammaproteobacteria (Cluster I) and Cluster III generally dominated the *nifH* DNA sequences in these studies. In addition, both spatial coupling (Deutsch et al., 2007) and decoupling (Knapp et al., 2016; Bonnet et al., 2017) of N₂ fixation and denitrification has been proposed. Collectively these studies suggest that NCNF is occurring in aphotic waters, and may correlate with oxygen availability, but the mechanisms by which oxygen and possibly other factors regulate NCNF in aphotic waters remain poorly known.

THE POTENTIAL ROLE OF DISSOLVED ORGANIC MATTER (DOM) IN SUPPORTING NCNF

Organic particles may provide a site of low oxygen and a source of DOM that both can benefit specific bacterial attachment (Thiele et al., 2015; Dang and Lovell, 2016). If NCDs are heterotrophic (Riemann et al., 2010), overall DOM availability should play a role in controlling their growth and activity (Kirchman, 1990). Genomic and physiological analyses have confirmed that gamma- and alphaproteobacterial diazotrophs isolated from the Baltic Sea are heterotrophic and contain genes responsible for DOM metabolism (Bentzon-Tilia et al., 2015a). Evidence for DOM influence on NCDs was reported from the South Pacific Gyre (SPG) photic layers (Halm et al., 2012) and in the Mediterranean Sea (Rahav et al., 2016), where DOM originating from primary production was suggested to impact N₂ fixation. Most of the SPG sequences were gammaproteobacterial, but represented different groups from those found in the ETSP (Fernandez et al., 2011; Turk-Kubo et al., 2014). Mesopelagic N₂ fixation rates had a positive correlation with relatively labile DOM (Benavides et al., 2015), and were enhanced upon the addition of sugars, amino acids, or transparent exopolymeric particles (Bonnet et al., 2013; Rahav et al., 2013a; Loescher et al., 2014; Benavides et al., 2015).

In the eastern Mediterranean Sea, N₂ fixation in the photic zone was uncoupled from primary production and correlated significantly and positively with bacterial production (Rahav et al., 2013b). However, even in coastal waters with high dissolved inorganic nutrient loads, organic carbon stimulated light and dark N₂ fixation in a community containing cyanobacterial and NCD *nifH* phylotypes (Rahav et al., 2016). Saharan dust addition (serving potentially as a source of trace elements, nutrients, and/or DOM) enhanced N₂ fixation and both NCD (Gamma A) and cyanobacterial diazotroph abundances in the North Atlantic (Langlois et al., 2012). Gamma A abundances were also enhanced by addition of sugars in the South Pacific (Moisander et al., 2012). Many cyanobacteria, including diazotrophs, take up organic forms of carbon (Hietanen et al., 2002; Church et al., 2004; Moisander et al., 2012; Benavides et al., 2017) thus rates of N₂ fixation in mixed communities, measured after DOM amendments, may reflect responses of both cyanobacterial and NCDs. Whether some marine NCDs also use light as an energy source remains to be demonstrated (Bombar et al., 2016).

CURRENT AND FUTURE CHALLENGES

The data synthesis here shows that while N₂ fixation rates have been reported by several studies in waters dominated by NCD *nifH* sequences (Table 1), most studies of NCDs did not measure transcription. The *nifH* DNA sequences often do not appear as transcripts in the same samples, suggesting that some of the organisms are not active (Moisander et al., 2006; Short and Zehr, 2007; Yogeve et al., 2011; Halm et al., 2012; Severin et al., 2015), thus it would be misleading to use *nifH* gene (DNA) data as proof for NCNF. Despite the common

detection of cyanobacterial and NCD *nifH* in DNA sequence libraries and, at times, in transcripts, only a few studies from surface layers reported the absence of cyanobacterial diazotrophs when N₂ fixation rates were detected (Table 1). Due to various methodological constraints, proving the absence of low numbers of cyanobacterial cells in a sample is difficult, if not impossible, yet such low abundance may be sufficient to result in detectable N₂ fixation rates. Aphotic waters may be considered a good case study for NCNF, as photoautotrophic N₂ fixation in these waters is conceivably absent. However, the common detection of diazotrophic cyanobacteria in aphotic layers (Hamersley et al., 2011; Farnelid et al., 2013; Benavides et al., 2015), possibly due to either settling material that could be viable upon experimental incubations (Agusti et al., 2015), or caused by contamination during sampling, complicate attributing measured aphotic N₂ fixation rates to NCDs alone. Overall, the measured rates of *in situ* aphotic N₂ fixation are higher than the parallel abundance and transcript numbers of NCDs would potentially support. Moreover, the reported cell-specific rates of NCDs are under debate (Turk-Kubo et al., 2014; Benavides et al., 2015; Bentzon-Tilia et al., 2015a; Gradoville et al., 2017).

Various factors of the ¹⁵N₂ method, such as uncertainties in the ¹⁵N-labeling step, influence N₂ fixation rate determination (Mohr et al., 2010; Grosskopf et al., 2012; Wilson et al., 2012). In addition, commercially available ¹⁵N₂ gases are at times contaminated with substrates other than N₂ (Dabundo et al., 2014), which could lead to false positive NCNF rates. An additional source of uncertainty that must be considered when assessing minimum quantifiable N₂ fixation rates is the concentration and isotopic composition of particulate organic N (PON), which at typical concentrations requires large volumes in incubations (usually >4 L) to constrain rates in deep waters. While N₂ fixation rates per volume are most informative for budgetary calculations, rates normalized to PON concentration would provide an additional measure for comparing rates across studies. Further, variability of the natural δ¹⁵N background of PON, changes in the δ¹⁵N of PON that may occur over the incubation period or due to substrate additions, and δ¹⁵N of the N₂ pool, should be considered (Montoya et al., 1996; Gradoville et al., 2017). These sources of uncertainty are not routinely reported and detection limits are infrequently calculated. Taking into account these sources of error, Gradoville et al. (2017) estimated the minimum quantifiable N₂ fixation rate in their study at ~0.4 nmol L⁻¹ d⁻¹ which is higher than many reported rates of NCNF (Table 1).

To our knowledge, direct field measurements of N₂ fixation per cell are currently lacking for marine NCDs; such measurements would be important in assessing their contribution to N₂ fixation rates, along with other efforts to separate signals of NCNF and cyanobacterial N₂ fixation (see also Bombar et al., 2016). High NCD *nifH* diversity renders identification and quantification of biogeochemically significant individual groups challenging. The combination of *in situ* hybridization approaches using halogenated probes together with single-cell mass spectrometry (nanoscale secondary ion mass spectrometry; nanoSIMS) has recently advanced quantification of single-cell N₂ fixation rates in UCYN-A

(Thompson et al., 2012; Krupke et al., 2013). Similar approaches could provide insights into the NCNF in marine environments. Stable isotope probing and isotope microarrays could lead to valuable future insights (Seyler et al., 2014; Arandia-Gorostidi et al., 2017).

In describing the communities, sequencing depth and primer specificity influence what portion of the NCD community is detected. In addition, the detection limit for diazotroph transcripts or genes and the detection limit for N₂ fixation are not necessarily equal. The quantification limit of quantitative PCR is often on the order of 10² *nifH* gene copies L⁻¹ (and often unreported), which could potentially result in false negatives for cyanobacterial diazotrophs, and subsequently, lead to false positive NCNF rates. Using small water volumes when abundance of cyanobacterial diazotrophs is low would increase the chances of reporting false negatives for these organisms. On the other hand, metabolic rate measurements of microbial samples brought to the surface from aphotic depths may be underestimated (Tamburini et al., 2013), making both rate and transcription analyses of deep communities challenging.

CONCLUSIONS

Our compiled analysis of data illustrates that low N₂ fixation rates were reported from marine environments where NCD abundance was high and cyanobacterial diazotrophs were low or undetected. NCD *nifH* sequences show a wider geographical and depth distribution in pelagic environments overall than their cyanobacterial counterparts. The emerging data suggest that the NCD communities are diverse but only a few groups have been identified that appear in several studies (Farnelid et al., 2011;

Turk-Kubo et al., 2014). Studies on NCD *nifH* transcripts in aphotic layers are scarce or missing for many areas of the oceans.

We currently lack a fundamental understanding of the key players and environmental regulation of NCNF in the ocean. Ecophysiological data of the organisms contributing to these rates are still preliminary and incomplete. Moreover, the measured rates in most environments are generally “snapshots” determined during a cruise/sampling foray and have not been examined over seasonal and annual cycles. In addition, several methodological concerns complicate interpretation of N₂ fixation rate data. Modeling the significance of NCNF in marine N₂ fixation remains a challenge due to these various constraints. If these rates are confirmed, however, NCNF could contribute significantly to new N inputs to the ocean. How factors such as temperature, DIN, oxygen, trace elements, and hydrostatic pressure drive the metabolic capacities and adaptations of NCNF are currently only partially revealed. The actual activities and taxon specific roles of marine NCDs remain enigmatic at present, as do their contributions to regional and global N₂ fixation.

AUTHOR CONTRIBUTIONS

PM and LR designed the study, wrote the initial drafts, and compiled the majority of **Table 1**. All authors wrote sections of the manuscript and contributed to **Table 1**.

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REFERENCES

- Agusti, S., Gonzalez-Gordillo, J. I., Vaque, D., Estrada, M., Cerezo, M. I., Salazar, G., et al. (2015). Ubiquitous healthy diatoms in the deep sea confirm deep carbon injection by the biological pump. *Nat. Commun.* 6:7608. doi: 10.1038/ncomms8608
- Arandia-Gorostidi, N., Weber, P. K., Alonso-Saez, L., Moran, X. A. G., and Mayali, X. (2017). Elevated temperature increases carbon and nitrogen fluxes between phytoplankton and heterotrophic bacteria through physical attachment. *ISME J.* 11, 641–650. doi: 10.1038/ismej.2016.156
- Benavides, M., Berthelot, H., Duhamel, S., Raimbault, P., and Bonnet, S. (2017). Dissolved organic matter uptake by *Trichodesmium* in the Southwest Pacific. *Sci. Rep.* 7:41315. doi: 10.1038/srep41315
- Benavides, M., Bonnet, S., Hernandez, N., Martinez-Perez, A. M., Nieto-Cid, M., Alvarez-Salgado, X. A., et al. (2016). Basin-wide N₂ fixation in the deep waters of the Mediterranean Sea. *Glob. Biogeochem. Cycles* 30, 952–961. doi: 10.1002/2015gb005326
- Benavides, M., Moisander, P. H., Berthelot, H., Dittmar, T., Grosso, O., and Bonnet, S. (2015). Mesopelagic N₂ fixation related to organic matter composition in the Solomon and Bismarck Seas (Southwest Pacific). *PLoS ONE* 10:e0143775. doi: 10.1371/journal.pone.0143775
- Bentzon-Tilia, M., Severin, I., Hansen, L. H., and Riemann, L. (2015a). Genomics and ecophysiology of heterotrophic nitrogen-fixing bacteria isolated from estuarine surface water. *MBio* 6:e00929. doi: 10.1128/mBio.00929-15
- Bentzon-Tilia, M., Traving, S. J., Mantikci, M., Knudsen-Leerbeck, H., Hansen, J. L., Stiig, M., et al. (2015b). Significant N₂ fixation by heterotrophs, photoheterotrophs and heterocystous cyanobacteria in two temperate estuaries. *ISME J.* 9, 273–285. doi: 10.1038/ismej.2014.119
- Berman-Frank, I., Chen, Y.-B., Gao, Y., Fennel, K., Follows, M. J., Milligan, A. J., et al. (2008). “Feedbacks between the nitrogen, carbon and oxygen cycles,” in *Nitrogen in the Marine Environment, 2nd Edn*, eds D. G. Capone, D. A. Bronk, M. R. Mulholland, and E. J. Carpenter (Burlington, MA: Elsevier, Inc.), 1537–1563.
- Berthelot, H., Benavides, M., Moisander, P. H., Grosso, O., and Bonnet, S. (2017). High nitrogen fixation rates in the particulate and dissolved pools in the Western Tropical Pacific (Solomon and Bismarck Seas). *Geophys. Res. Lett.* doi: 10.1002/2017GL073856. [Epub ahead of print].
- Bird, C., and Wyman, M. (2013). Transcriptionally active heterotrophic diazotrophs are widespread in the upper water column of the Arabian Sea. *FEMS Microb. Ecol.* 84, 189–200. doi: 10.1111/1574-6941.12049
- Bird, C., Martinez, J. M., O'Donnell, A. G., and Wyman, M. (2005). Spatial distribution and transcriptional activity of an uncultured clade of planktonic diazotrophic gamma-proteobacteria in the Arabian Sea. *Appl. Environ. Microbiol.* 71, 2079–2085. doi: 10.1128/AEM.71.4.2079-2085.2005
- Blais, M., Tremblay, J.-E., Jungblut, A. D., Gagnon, J., Martin, J., Thaler, M., et al. (2012). Nitrogen fixation and identification of potential diazotrophs in the Canadian Arctic. *Global Biogeochem. Cycles* 26:GB3022. doi: 10.1029/2011GB004096
- Bombar, D., Paerl, R. W., and Riemann, L. (2016). Marine non-cyanobacterial diazotrophs: moving beyond molecular detection. *Trends Microbiol.* 24, 916–927. doi: 10.1016/j.tim.2016.07.002
- Bonnet, S., Caffin, M., Berthelot, H., and Moutin, T. (2017). Hot spot of N₂ fixation in the western tropical South Pacific pleads for a spatial decoupling between

- N₂ fixation and denitrification. *Proc. Natl. Acad. Sci. U.S.A.* 114, E2800–E2801. doi: 10.1073/pnas.1619514114
- Bonnet, S., Dekaezemaeker, J., Turk-Kubo, K. A., Moutin, T., Hamersley, R. M., Grosso, O., et al. (2013). Aphotic N₂ fixation in the Eastern Tropical South Pacific Ocean. *PLoS ONE* 8:e81265. doi: 10.1371/journal.pone.0081265
- Bonnet, S., Rodier, M., Turk, K., Germineaud, C., Menkes, C., Ganachaud, A., et al. (2015). Contrasted geographical distribution of N₂ fixation rates and *nifH* phylotypes in the Coral and Solomon Seas (South-Western Pacific) during austral winter conditions. *Global Biogeochem. Cycles* 29, 1874–1892. doi: 10.1002/2015GB005117
- Brown, S. M., and Jenkins, B. D. (2014). Profiling gene expression to distinguish the likely active diazotrophs from a sea of genetic potential in marine sediments. *Environ. Microbiol.* 16, 3128–3142. doi: 10.1111/1462-2920.12403
- Capone, D. G., Burns, J. A., Montoya, J. P., Subramaniam, A., Mahaffey, C., Gunderson, T., et al. (2005). Nitrogen fixation by *Trichodesmium* spp.: an important source of new nitrogen to the tropical and subtropical North Atlantic Ocean. *Global Biogeochem. Cycles* 19:GB2024. doi: 10.1029/2004GB002331
- Chien, Y.-T., and Zinder, S. H. (1996). Cloning, functional organization, transcript studies, and phylogenetic analysis of the complete nitrogenase structural genes (*nifHDK2*) and associated genes in the archaeon *Methanosarcina barkeri* 227. *J. Bacteriol.* 178, 143–148. doi: 10.1128/jb.178.1.143-148.1996
- Church, M. J., Ducklow, H. W., and Karl, D. A. (2004). Light dependence of H-3 leucine incorporation in the oligotrophic North Pacific ocean. *Appl. Environ. Microbiol.* 70, 4079–4087. doi: 10.1128/AEM.70.7.4079-4087.2004
- Church, M. J., Jenkins, B. D., Karl, D. M., and Zehr, J. P. (2005a). Vertical distributions of nitrogen-fixing phylotypes at Stn ALOHA in the oligotrophic North Pacific Ocean. *Aquat. Microb. Ecol.* 38, 3–14. doi: 10.3354/ame038003
- Church, M. J., Short, C. M., Jenkins, B. D., Karl, D. M., and Zehr, J. P. (2005b). Temporal patterns of nitrogenase gene (*nifH*) expression in the oligotrophic North Pacific Ocean. *Appl. Environ. Microbiol.* 71, 5362–5370. doi: 10.1128/AEM.71.9.5362-5370.2005
- Dabundo, R., Lehmann, M. F., Treibergs, L., Tobias, C. R., Altabet, M. A., Moisander, P. H., et al. (2014). The contamination of commercial ¹⁵N₂ gas stocks with ¹⁵N-labeled nitrate and ammonium and consequences for nitrogen fixation measurements. *PLoS ONE* 9:e110335. doi: 10.1371/journal.pone.0110335
- Dang, H., and Lovell, C. R. (2016). Microbial surface colonization and biofilm development in marine environments. *Microbiol. Mol. Biol. Rev.* 80, 91–138. doi: 10.1128/MMBR.00037-15
- Dekaezemaeker, J., Bonnet, S., Grosso, O., Moutin, T., Bressac, M., and Capone, D. G. (2013). Evidence of active dinitrogen fixation in surface waters of the eastern tropical South Pacific during El Niño and La Niña events and evaluation of its potential nutrient controls. *Global Biogeochem. Cycles* 27, 768–779. doi: 10.1002/gbc.20063
- Dekas, A. E., Chadwick, G. L., Bowles, M. W., Joye, S. B., and Orphan, V. J. (2014). Spatial distribution of nitrogen fixation in methane seep sediment and the role of the ANME archaea. *Environ. Microbiol.* 16, 3012–3029. doi: 10.1111/1462-2920.12247
- Dekas, A. E., Poretsky, R. S., and Orphan, V. J. (2009). Deep-sea archaea fix and share nitrogen in methane-consuming microbial consortia. *Science* 326, 422–426. doi: 10.1126/science.1178223
- Delmont, T. O., Quince, C., Shaiber, A., Esen, O. C., Lee, S. T. M., Lucker, S., et al. (2017). Nitrogen fixing populations of Planctomycetes and Proteobacteria are abundant in the surface ocean. *BioRxiv*. doi: 10.1101/129791
- Deutscher, C., Sarmiento, J. L., Sigman, D. M., Gruber, N., and Dunne, J. P. (2007). Spatial coupling of nitrogen inputs and losses in the ocean. *Nature* 445, 163–167. doi: 10.1038/nature05392
- Falcon, L. I., Carpenter, E. J., Cipriano, F., Bergman, B., and Capone, D. G. (2004). N₂ fixation by unicellular bacterioplankton from the Atlantic and Pacific oceans: phylogeny and *in situ* rates. *Appl. Environ. Microbiol.* 70, 765–770. doi: 10.1128/AEM.70.2.765-770.2004
- Farnelid, H., Andersson, A. F., Bertilsson, S., Abu Al-Soud, W., Hansen, L. H., Sørensen, S., et al. (2011). Nitrogenase gene amplicons from global marine surface waters are dominated by genes of non-cyanobacteria. *PLoS ONE* 6:e19223. doi: 10.1371/journal.pone.0019223
- Farnelid, H., Bentzon-Tilia, M., Andersson, A. F., Bertilsson, S., Jost, G., Labrenz, M., et al. (2013). Active nitrogen-fixing heterotrophic bacteria at and below the chemocline of the central Baltic Sea. *ISME J.* 7, 1413–1423. doi: 10.1038/ismej.2013.26
- Fay, P. (1992). Oxygen relations of nitrogen fixation in cyanobacteria. *Microbiol. Rev.* 56, 340–373.
- Fernandez, C., Farias, L., and Ulloa, O. (2011). Nitrogen fixation in denitrified marine waters. *PLoS ONE* 6:e20539. doi: 10.1371/journal.pone.0020539
- Foster, R. A., Paytan, A., and Zehr, J. P. (2009). Seasonality of N₂ fixation and *nifH* gene diversity in the Gulf of Aqaba (Red Sea). *Limnol. Oceanogr.* 54, 219–233. doi: 10.4319/lo.2009.54.1.0219
- Gradoville, M. R., Bombar, D., Crump, B., Letelier, R. M., Zehr, J. P., and White, A. E. (2017). Diversity and activity of nitrogen-fixing communities across ocean basins. *Limnol. Oceanogr.* doi: 10.1002/lno.10542. [Epub ahead of print].
- Grosskopf, T., Mohr, W., Baustian, T., Schunck, H., Gill, D., Kuypers, M. M. M., et al. (2012). Doubling of marine dinitrogen-fixation rates based on direct measurements. *Nature* 488, 361–364. doi: 10.1038/nature11338
- Halm, H., Lam, P., Ferdelman, T. G., Lavik, G., Dittmar, T., LaRoche, J., et al. (2012). Heterotrophic organisms dominate nitrogen fixation in the South Pacific Gyre. *ISME J.* 6, 1238–1249. doi: 10.1038/ismej.2011.182
- Hamersley, M. R., Turk, K. A., Leinweber, A., Gruber, N., Zehr, J. P., Gunderson, T., et al. (2011). Nitrogen fixation within the water column associated with two hypoxic basins in the Southern California Bight. *Aquat. Microb. Ecol.* 63, 193–205. doi: 10.3354/ame01494
- Hayat, R., Ali, S., Amara, U., Khalid, R., and Ahmed, I. (2010). Soil beneficial bacteria and their role in plant growth promotion: a review. *Ann. Microbiol.* 60, 579–598. doi: 10.1007/s13213-010-0117-1
- Herridge, D. F., Peoples, M. B., and Boddey, R. M. (2008). Global inputs of biological nitrogen fixation in agricultural systems. *Plant Soil* 311, 1–18. doi: 10.1007/s11104-008-9668-3
- Hewson, I., Moisander, P. H., Achilles, K. M., Carlson, C. A., Jenkins, B. D., Mondragon, E. A., et al. (2007). Characteristics of diazotrophs in surface to abyssopelagic waters of the Sargasso Sea. *Aquat. Microb. Ecol.* 46, 15–30. doi: 10.3354/ame046015
- Hietanen, S., Lehtimäki, J. M., Tuominen, L., Sivonen, K., and Kuparinen, J. (2002). *Nodularia* spp. (Cyanobacteria) incorporate leucine but not thymidine: importance for bacterial-production measurements. *Aquat. Microb. Ecol.* 28, 99–104. doi: 10.3354/ame028099
- Hood, R. R., Coles, V. J., and Capone, D. G. (2004). Modeling the distribution of *Trichodesmium* and nitrogen fixation in the Atlantic Ocean. *J. Geophys. Res.* 109:C06006. doi: 10.1029/2002JC001753
- Jayakumar, A., Al-Rshaidat, M. M. D., Ward, B. B., and Mulholland, M. R. (2012). Diversity, distribution, and expression of diazotroph *nifH* genes in oxygen-deficient waters of the Arabian Sea. *FEMS Microb. Ecol.* 82, 597–606. doi: 10.1111/j.1574-6941.2012.01430.x
- Karl, D., Michaels, A., Bergman, B., Capone, D., Carpenter, E., Letelier, R., et al. (2002). Dinitrogen fixation in the world's oceans. *Biogeochemistry* 57–58, 47–98. doi: 10.1023/A:1015798105851
- Kirchman, D. L. (1990). Limitation of bacterial growth by dissolved organic matter in the subarctic Pacific. *Mar. Ecol. Prog. Ser.* 62, 47–54. doi: 10.3354/meps062047
- Kirshtein, J. D., Paerl, H. W., and Zehr, J. P. (1991). Amplification, cloning, and sequencing of a *nifH* segment from aquatic microorganisms and natural communities. *Appl. Environ. Microbiol.* 57, 2645–2650.
- Knapp, A. N. (2012). The sensitivity of marine N₂ fixation to dissolved inorganic nitrogen. *Front. Microbiol.* 3:374. doi: 10.3389/fmicb.2012.00374
- Knapp, A. N., Casciotti, K. L., Berelson, W. M., Prokopenko, M. G., and Capone, D. G. (2016). Low rates of nitrogen fixation in eastern tropical South Pacific surface waters. *Proc. Natl. Acad. Sci. U.S.A.* 113, 4398–4403. doi: 10.1073/pnas.1515641113
- Krupke, A., Musat, N., LaRoche, J., Mohr, W., Fuchs, B. M., Amann, R. L., et al. (2013). *In situ* identification and N₂ and C fixation rates of uncultivated cyanobacteria populations. *Syst. Appl. Microbiol.* 36, 259–271. doi: 10.1016/j.syapm.2013.02.002
- Langlois, R. J., LaRoche, J., and Raab, P. A. (2005). Diazotrophic diversity and distribution in the tropical and subtropical Atlantic ocean. *Appl. Environ. Microbiol.* 71, 7910–7919. doi: 10.1128/AEM.71.12.7910-7919.2005
- Langlois, R. J., Mills, M. M., Ridame, C., Croot, P., and LaRoche, J. (2012). Diazotrophic bacteria respond to Saharan dust additions. *Mar. Ecol. Prog. Ser.* 470, 1–14. doi: 10.3354/meps10109

- Langlois, R., Grokopf, T., Mills, M., Takeda, S., and LaRoche, J. (2015). Widespread distribution and expression of Gamma A (UMB), an uncultured, diazotrophic, gamma-Proteobacterial *nifH* Phylotype. *PLoS ONE* 10:e0128912. doi: 10.1371/journal.pone.0128912
- Loescher, C. R., Grosskopf, T., Desai, F. D., Gill, D., Schunck, H., Croot, P. L., et al. (2014). Facets of diazotrophy in the oxygen minimum zone waters off Peru. *ISME J.* 8, 2180–2192. doi: 10.1038/ismej.2014.71
- Mahaffey, C., Michaels, A. F., and Capone, D. G. (2005). The conundrum of marine N-2 fixation. *Am. J. Sci.* 305, 546–595. doi: 10.2475/ajs.305.6-8.546
- Man-Aharonovich, D., Kress, N., Bar Zeev, E., Berman-Frank, I., and Beja, O. (2007). Molecular ecology of *nifH* genes and transcripts in the eastern Mediterranean Sea. *Environ. Microbiol.* 9, 2354–2363. doi: 10.1111/j.1462-2920.2007.01353.x
- Mehta, M. P., and Baross, J. A. (2006). Nitrogen fixation at 92 degrees C by a hydrothermal vent archaeon. *Science* 314, 1783–1786. doi: 10.1126/science.1134772
- Messer, L. F., Mahaffey, C., Robinson, C. M., Jeffries, T. C., Baker, K. G., Isaksson, J. B., et al. (2016). High levels of heterogeneity in diazotroph diversity and activity within a putative hotspot for marine nitrogen fixation. *ISME J.* 10, 1499–1513. doi: 10.1038/ismej.2015.205
- Mohr, W., Grosskopf, T., Wallace, D. W. R., and LaRoche, J. (2010). Methodological underestimation of oceanic nitrogen fixation rates. *PLoS ONE* 5:e12583. doi: 10.1371/journal.pone.0012583
- Moisander, P. H., Beinart, R. A., Voss, M., and Zehr, J. P. (2008). Diversity and abundance of diazotrophic microorganisms in the South China Sea during intermonsoon. *ISME J.* 2, 954–967. doi: 10.1038/ismej.2008.51
- Moisander, P. H., Serros, T., Paerl, R. W., Beinart, R. A., and Zehr, J. P. (2014). Gammaproteobacterial diazotrophs and *nifH* gene expression in surface waters of the South Pacific Ocean. *ISME J.* 8, 1962–1973. doi: 10.1038/ismej.2014.49
- Moisander, P. H., Shiue, L., Steward, G. F., Jenkins, B. D., Bebout, B. M., and Zehr, J. P. (2006). Application of a *nifH* oligonucleotide microarray for profiling diversity of N-2-fixing microorganisms in marine microbial mats. *Environ. Microbiol.* 8, 1721–1735. doi: 10.1111/j.1462-2920.2006.01108.x
- Moisander, P. H., Zhang, R., Boyle, E. A., Hewson, I., Montoya, J. P., and Zehr, J. P. (2012). Analogous nutrient limitations in unicellular diazotrophs and *Prochlorococcus* in the South Pacific Ocean. *ISME J.* 6, 733–744. doi: 10.1038/ismej.2011.152
- Montoya, J. P., Voss, M., Kähler, P., and Capone, D. G. (1996). A simple, high-precision, high-sensitivity tracer assay for N₂ fixation. *Appl. Environ. Microbiol.* 62, 986–993.
- Mortenson, L. E., and Thorneley, R. N. F. (1979). Structure and function of nitrogenase. *Ann. Rev. Biochem.* 48, 387–418. doi: 10.1146/annurev.bi.48.070179.002131
- Mulholland, M. R., Bernhardt, P. W., Heil, C. A., Bronk, D. A., and O'Neil, J. M. (2006). Nitrogen fixation and release of fixed nitrogen by *Trichodesmium* spp. in the Gulf of Mexico. *Limnol. Oceanogr.* 51, 1762–1776. doi: 10.4319/lo.2006.51.4.1762
- Postgate, J. (1998). *Nitrogen Fixation, 3rd Edn.* New York, NY: Cambridge University Press.
- Rahav, E., Bar-Zeev, E., Ohayon, S., Elifantz, H., Belkin, N., Herut, B., et al. (2013a). Dinitrogen fixation in aphotic oxygenated marine environments. *Front. Microbiol.* 4:227. doi: 10.3389/fmicb.2013.00227
- Rahav, E., Giannetto, M. J., and Bar-Zeev, E. (2016). Contribution of mono and polysaccharides to heterotrophic N₂ fixation at the eastern Mediterranean coastline. *Sci. Rep.* 6:27858. doi: 10.1038/srep27858
- Rahav, E., Herut, B., Stambler, N., Bar-Zeev, E., Mulholland, M. R., and Berman-Frank, I. (2013b). Uncoupling between dinitrogen fixation and primary productivity in the eastern Mediterranean Sea. *J. Geophys. Res.* 118, 195–202. doi: 10.1002/jgrg.20023
- Riemann, L., Farnelid, H., and Steward, G. F. (2010). Nitrogenase genes in non-cyanobacterial plankton: prevalence, diversity, and regulation in marine waters. *Aquat. Microb. Ecol.* 61, 235–247. doi: 10.3354/ame01431
- Severin, I., Bentzon-Tilia, M., Moisander, P. H., and Riemann, L. (2015). Nitrogenase expression in estuarine bacterioplankton influenced by organic carbon and availability of oxygen. *FEMS Microb. Lett.* 326:fnv105. doi: 10.1093/femsle/fnv105
- Seyler, L. M., McGuinness, L. M., and Kerkhof, L. J. (2014). Crenarchaeal heterotrophy in salt marsh sediments. *ISME J.* 8, 1534–1543. doi: 10.1038/ismej.2014.15
- Shiozaki, T., Ijichi, M., Kodama, T., Takeda, S., and Furuya, K. (2014). Heterotrophic bacteria as major nitrogen fixers in the euphotic zone of the Indian Ocean. *Global Biogeochem. Cycles* 28, 1096–1110. doi: 10.1002/2014GB004886
- Short, S. M., and Zehr, J. P. (2007). Nitrogenase gene expression in the Chesapeake Bay Estuary. *Environ. Microbiol.* 9, 1591–1596. doi: 10.1111/j.1462-2920.2007.01258.x
- Sohm, J. A., Hilton, J. A., Noble, A. E., Zehr, J. P., Saito, M. A., and Webb, E. A. (2011). Nitrogen fixation in the South Atlantic Gyre and the Benguela Upwelling System. *Geophys. Res. Lett.* 38:L16608. doi: 10.1029/2011GL048315
- Tamburini, C., Boutrif, M., Garel, M., Colwell, R. R., and Deming, J. W. (2013). Prokaryotic responses to hydrostatic pressure in the ocean - a review. *Environ. Microbiol.* 15, 1262–1274. doi: 10.1111/1462-2920.12084
- Thiele, S., Fuchs, B. M., Amann, R., and Iversen, M. H. (2015). Colonization in the photic zone and subsequent changes during sinking determine bacterial community composition in marine snow. *Appl. Environ. Microbiol.* 81, 1463–1471. doi: 10.1128/AEM.02570-14
- Thompson, A. W., Foster, R. A., Krupke, A., Carter, B. J., Musat, N., Vault, D., et al. (2012). Unicellular cyanobacterium symbiotic with a single-celled eukaryotic alga. *Science* 337, 1546–1550. doi: 10.1126/science.1222700
- Turk-Kubo, K. A., Karamchandani, M., Capone, D. G., and Zehr, J. P. (2014). The paradox of marine heterotrophic nitrogen fixation: abundances of heterotrophic diazotrophs do not account for nitrogen fixation rates in the Eastern Tropical South Pacific. *Environ. Microbiol.* 16, 3095–3114. doi: 10.1111/1462-2920.12346
- Wilson, S. T., Boettjer, D., Church, M. J., and Karl, D. M. (2012). Comparative assessment of nitrogen fixation methodologies, conducted in the oligotrophic North Pacific Ocean. *Appl. Environ. Microbiol.* 78, 6516–6523. doi: 10.1128/AEM.01146-12
- Yogev, T., Rahav, E., Bar-Zeev, E., Man-Aharonovich, D., Stambler, N., Kress, N., et al. (2011). Is nitrogen fixation significant in the Levantine Basin, East Mediterranean Sea? *Environ. Microbiol.* 13, 854–871. doi: 10.1111/j.1462-2920.2010.02402.x
- Zehr, J. P. (2011). Nitrogen fixation by marine cyanobacteria. *Trends Microbiol.* 19, 162–173. doi: 10.1016/j.tim.2010.12.004
- Zehr, J. P., Jenkins, B. D., Short, S. M., and Steward, G. F. (2003). Nitrogenase gene diversity and microbial community structure: a cross-system comparison. *Environ. Microbiol.* 5, 539–554. doi: 10.1046/j.1462-2920.2003.00451.x
- Zehr, J. P., Mellon, M. T., and Zani, S. (1998). New nitrogen-fixing microorganisms detected in oligotrophic oceans by amplification of nitrogenase (*nifH*) genes. *Appl. Environ. Microbiol.* 64, 3444–3450.
- Zehr, J. P., Mellon, M., Braun, S., Litaker, W., Steppe, T., and Paerl, H. W. (1995). Diversity of heterotrophic nitrogen fixation genes in a marine cyanobacterial mat. *Appl. Environ. Microbiol.* 61, 2527–2532.
- Zehr, J. P., Harris, D., Dominic, B., and Salerno, J. (1997). Structural analysis of the *Trichodesmium* nitrogenase iron protein: implications for aerobic nitrogen fixation activity. *FEMS Microb. Lett.* 153, 303–309. doi: 10.1111/j.1574-6968.1997.tb12589.x

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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